

SUPPLEMENTATION WITH THREE DIFFERENT MACULAR CAROTENOID FORMULATIONS IN PATIENTS WITH EARLY AGE-RELATED MACULAR DEGENERATION

SARAH SABOUR-PICKETT, PhD,*†‡ STEPHEN BEATTY, MD, FRCOPHTH,†‡
EITHNE CONNOLLY, BSc,†‡ JAMES LOUGHMAN, PhD,*§ JIM STACK, PhD,† ALAN HOWARD, PhD,¶
RONALD KLEIN, MD, MPH,** BARBARA E. KLEIN, MD, MPH,** STACY M. MEUER, BSc,**
CHELSEA E. MYERS, MStat,** KWADWO O. AKUFFO, OD,† JOHN M. NOLAN, PhD†‡

Purpose: To investigate the impact of three different macular carotenoid formulations on macular pigment optical density and visual performance in subjects with early age-related macular degeneration.

Methods: Fifty-two subjects were supplemented and followed for 12 months, 17 of them were in intervention Group 1 (20 mg/day lutein and 2 mg/day zeaxanthin); 21 in Group 2 (10 mg/day *meso*-zeaxanthin, 10 mg/day lutein, and 2 mg/day zeaxanthin); and 14 in Group 3 (17 mg/day *meso*-zeaxanthin, 3 mg/day lutein, and 2 mg/day zeaxanthin). The macular pigment optical density was measured using customized heterochromatic flicker photometry, and visual function was assessed using corrected distance visual acuity and by letter contrast sensitivity.

Results: A statistically significant increase in the macular pigment optical density was observed at all measured eccentricities in Group 2 ($P \leq 0.005$) and in Group 3 ($P < 0.05$, for all), but only at 1.75° in Group 1 ($P = 0.018$). Statistically significant ($P < 0.05$) improvements in letter contrast sensitivity were seen at all spatial frequencies (except 1.2 cycles per degree) in Group 3, and at low spatial frequencies in Groups 1 and 2.

Conclusion: Augmentation of the macular pigment optical density across its spatial profile and enhancements in contrast sensitivity were best achieved after supplementation with a formulation containing high doses of *meso*-zeaxanthin in combination with lutein and zeaxanthin.

RETINA 0:1–10, 2014

The prevalence of age-related macular degeneration (AMD), the leading cause of blind registration in the developed world,¹ is rising because of increasing longevity.^{2,3} Although antivascular endothelial growth factor therapy has resulted in better outcomes for patients with neovascular AMD,⁴ this treatment is expensive and cumbersome to the patient and to the health care provider.

Investigators interested in exploring ways of preventing, delaying the onset, or retarding the progression of AMD have directed their attention toward the possible protective role of macular pigment (MP), a yellow-colored pigment that accumulates within the inner retinal layers at the macula⁵ and is

optically undetectable beyond 7° eccentricity.⁶ Macular pigment is composed of three carotenoids, lutein (L), zeaxanthin (Z), and *meso*-zeaxanthin (MZ).^{7,8} Macular pigment has generated interest in recent years because of its possible protective role for AMD, putatively attributable to its antioxidant properties and/or its pre-receptor filtration of damaging (short-wavelength) blue light, given that (photo-) oxidative retinal injury is known to be important in the pathogenesis of this condition.^{9,10}

Low levels of MP are associated with known risk factors for AMD, namely, increasing age, a positive family history of the condition, tobacco use and obesity, before the onset of disease.¹¹ Furthermore,

observational studies have shown that low levels of carotenoids in the diet^{12–16} and in the serum^{13,17–19} are associated with the risk of AMD. Importantly, MP augmentation has repeatedly been demonstrated after dietary modification and/or supplementation with its constituent carotenoids, in subjects with and without AMD.^{20–26}

Although L and Z concentrations in a variety of foodstuffs have been determined,^{27,28} the MZ composition of foodstuffs typical of a western diet has not been investigated satisfactorily,²⁸ although it has been identified in certain types of seafood.²⁹ Interestingly, MZ has been found, albeit in trace amounts, in the serum of subjects who have not been supplemented with this carotenoid.³⁰

There is consensus that MP plays an important role in visual performance. Many cross-sectional studies have shown a positive association between MP and measures of visual performance, including visual acuity, contrast sensitivity (CS), photostress recovery and glare disability (among others).^{31–35} It has also been shown that supplementation with the macular carotenoids improves parameters of visual function in patients afflicted with the early form of this condition.^{36–38} However, no study has yet investigated the impact of a formulation containing MZ on visual function in subjects with early AMD, or on the natural course of this condition.

Certain properties of MZ render this carotenoid of particular interest when investigating AMD prevention, or when studying the contribution that MP makes to visual performance and experience (in subjects with or without AMD), and these include: MZ is believed to be generated from L in the primate retina³⁹; MZ is the dominant carotenoid at the epicenter of the

macula⁴⁰; MZ seems to be the most powerful antioxidant of the macular carotenoids in the presence of the xanthophyll-binding proteins⁴¹; the presence of all three macular carotenoids is required if MP is to maximally exert its antioxidant effects⁴²; the presence of MZ facilitates a wider range of pre-receptor blue light filtration by MP.^{43,44} Interestingly, an atypical central dip in the spatial profile of MP, characterized by the lack of a central peak with a monotonic decline from the foveal center, is associated with risk for AMD.⁴⁵ It is reasonable to hypothesize that such atypical profiles may be attributable, at least in part, to a lack of MZ, and a consequential lack of MP at the site of dominance of this carotenoid (i.e., at the foveal center). Interestingly, supplementation with a formulation containing MZ has the ability, uniquely, to rebuild MP centrally and confer a typical central peak to its spatial profile.^{30,46}

This single-blind, randomized control trial was designed to compare the effect of three differing macular carotenoid formulations on MP enhancement, on visual performance, and on disease progression in subjects with early AMD.

Methods

Subjects and Study Design

This study was conducted at the Institute of Vision Research and Institute of Eye Surgery, Waterford, Ireland. The inclusion criteria were: early AMD (the presence of drusen and pigmentary changes) in at least 1 eye; corrected distance visual acuity of $\geq 6/12$ in the study eye. The exclusion criteria were: a recent history (within 3 months of baseline visit) of macular carotenoid supplementation; diabetes mellitus; any visually consequential ocular comorbidity. Ethics approval was granted by the Waterford Regional Hospital Ethics Committee, and written informed consent was secured from each subject. The research was conducted in accordance with the principles of the Declaration of Helsinki.

The Meso-zeaxanthin Ocular Supplementation Trial: Report 1 (trial registration number: ISRCTN60816411) is a randomized single-blind clinical trial of oral supplementation with 1 of 3 different interventions. Subjects were randomly assigned to one of three supplementation groups, as follows: Group 1: 20 mg L and 2 mg Z (Ultra Lutein, Natural Organics, Inc., Melville, NY); Group 2: 10 mg MZ, 10 mg L, and 2 mg Z (MacuHealth, [MacuHealth LLC, Birmingham, MI], or Macushield in non-North American territories); Group 3: 17 mg MZ, 3 mg L, and 2 mg Z (prepared especially for this trial by Industrial Organica, Monterrey, Mexico). Subjects were

From the *Department of Optometry, School of Physics, Dublin Institute of Technology, Dublin, Ireland; †Macular Pigment Research Group, Waterford Institute of Technology, Waterford, Ireland; ‡Institute of Eye Surgery and Institute of Vision Research, Whitfield Clinic, Waterford, Ireland; §African Vision Research Institute, Faculty of Health Sciences, University of KwaZulu-Natal, Durban, South Africa; ¶Howard Foundation, Cambridge, United Kingdom; and **Department of Ophthalmology and Visual Sciences, University of Wisconsin, Madison, Wisconsin.

This study was conducted at the Institute of Vision Research and Institute of Eye Surgery, Whitfield Clinic, Waterford, Ireland.

Supported by a grant from The Howard Foundation, Cambridge CB22 5LA, United Kingdom. The principal investigator (J.M.N.) is currently funded by the European Research Council.

J. Loughman, J. M. Nolan, and S. Beatty do consultancy work for Nutraceutical companies in a personal capacity. J. M. Nolan and S. Beatty are directors of Nutrasight Consultancy Limited. A. Howard is a Chairman of the Howard Foundation, a foundation that supports research in the field of nutrition and health. The other authors have no conflicting interests to disclose.

Reprint requests: John M. Nolan, PhD, Macular Pigment Research Group, Carriganore House, WIT, West Campus, Waterford, Ireland; e-mail: jmnolan@wit.ie

required to consume one tablet daily with a meal. Study visits were carried out at baseline and at 12 months. A demographic, medical, ophthalmic, and lifestyle case history was obtained for each patient at baseline.

Macular Pigment Optical Density

Each subject's MP spatial profile was obtained with the Macular Densitometer, using a methodology that has been slightly modified from that developed by Wooten et al.⁴⁷ A detailed description of this protocol has been previously described.^{48,49}

Visual Performance

Corrected distance visual acuity was measured for the study eye monocularly using the Early Treatment Diabetic Retinopathy Study logMAR chart (Test Chart 2000 PRO; Thomson Software Solutions, Hertfordshire, England, United Kingdom), with the room lights on. Contrast sensitivity was also assessed using the logMAR chart at 5 different spatial frequencies (1.2, 2.4, 6.0, 9.6, and 15.15 cycles per degree). For a given spatial frequency, subjects were asked to read out the letters while fixating on the chart at a distance of 6 m. The letter set was randomized during the test at each change of contrast. The percentage contrast of letter optotypes was reduced in 0.15 logCS steps until the lowest contrast value at which subjects see at least 3 letters was reached. The test is then repeated for the other spatial frequencies. Each letter has a nominal logCS value of 0.03. Missed letters at any contrast level are noted. The resultant logCS value for the subject at a particular spatial frequency is calculated by adding any extra letter(s) and/or subtracting missed letters from best logCS value corresponding to the lowest percentage contrast.

Morphologic Assessment

Subjects recruited into the study had early AMD. To establish AMD status, color stereoscopic 30° fundus photographs were obtained using a Zeiss VisuCam (Carl Zeiss Meditec AG, Jena, Germany) and were graded at the Ocular Epidemiology Reading Center at the University of Wisconsin, Madison, WI, using a modified version of the Wisconsin Age-Related Maculopathy Grading System and based on the 11-step AREDS grading scale.^{50,51} For the purposes of this study, a change of two or more steps along the AREDS severity scale was defined as being clinically meaningful.⁵²

Clinical Pathology Analysis

Clinical pathology analysis was performed by Biomnis Laboratories (Dublin, Ireland) to test for changes in renal and liver function, lipid profile, hematologic profile, and inflammation markers at baseline and after the 12-month supplementation period. A detailed description of the protocol has been previously described by our group.⁵³

Statistical Analysis

Statistical analysis was performed using the software package PASW Statistics 18.0 (IBM Corp, Somers, NY). Power and sample size calculations were obtained using PASS 2008 (NCSS, LLC, Kaysville, UT). A priori statistical methodology was not used in this exploratory study.

Baseline differences between intervention groups were assessed using analysis of variance and contingency table analysis, as appropriate. Baseline and 12-month visit measures, within each treatment group, were compared using the paired-samples *t*-test; between-group comparisons, using analysis of variance, would have lacked statistical power because of the relatively small sample size in this study. The change in AMD-severity grade between the three intervention groups was assessed using the Pearson chi-square test for contingency tables. The 5% level of significance was used throughout.

For the paired *t*-test analyses of changes in MP optical density (MPOD) and CS (reported in Tables 3 and 4), power calculations were based on a "large" effect size of 0.8 standard deviations (as suggested by Cohen⁵⁴), and on the smallest of the group sizes (Group 3, *n* = 14); this study was not powered to detect smaller effect sizes, as per Cohen's definitions.⁵⁴ With the usual assumptions (5% level of significance, 2-tailed test), the power is 0.79 for the Group 3 investigations and higher than this for the other two groups. For the contingency table analysis designed to detect changes on the AMD-severity scale (reported in Table 4), we also used a "large" effect size (*W* = 0.5 using Cohen's classification) and, in addition, combined some adjacent columns; in this case, the power is 0.78.

Results

Baseline Analysis

Sixty-seven eyes (of 67 subjects) were recruited into this study. Eight subjects discontinued for personal reasons, 3 were not available to attend for the 12-month visit, 2 discontinued for health reasons (deemed to be unrelated to intervention), 1 had cataract surgery

on the study eye before the 12-month visit, and 1 patient developed neovascular AMD and did not re-attend, leaving 52 subjects with complete data sets for the 12-month analyses: 17 in Group 1, 21 in Group 2, and 14 in Group 3. Baseline demographic, lifestyle, anthropometric, and visual data for the remaining 52 subjects are presented in Table 1. Of note, there was no significant difference between the groups in any baseline data variables (including AMD severity, data not presented).

Longitudinal Analysis

Values for MPOD at each eccentricity, at baseline, and 12 months are summarized in Table 2.

Letter CS at baseline and 12 months, for each of the 5 spatial frequencies, is summarized in Table 3. Graphical representations of letter CS at baseline and at 12 months, for the 3 intervention groups,

and for each spatial frequency, are displayed in Figure 1, A–C.

The proportion of subjects in each intervention group exhibiting a change in severity scale grade of two or more, considered clinically meaningful for the purpose of this study,⁵² was studied (see Table 4). Seventy-nine percent of subjects exhibited no clinically meaningful change in AMD severity grade between baseline and 12 months, with 11% exhibiting deterioration and 10% exhibiting an improvement. There was no statistically significant difference between treatment groups in terms of change in the AMD severity scale ($P = 0.455$, Pearson chi-square test). The 24- and 36-month data will further inform this important analysis.

Clinical pathology analysis results are reported in Table 5. Of note, 2 variables in Group 1, 2 variables in Group 2, and 2 variables in Group 3 demonstrated statistically significant changes from baseline (in both positive and negative directions). All variables,

Table 1. Baseline Demographic, Lifestyle, Anthropometric, and Visual Data

	Entire Group, n (%)	Group 1, n (%)	Group 2, n (%)	Group 3, n (%)	Significance
Gender					
Male	18 (35)	5 (29)	8 (38)	5 (36)	0.851
Female	34 (65)	12 (71)	13 (62)	9 (64)	
Laterality					
Right	33 (63)	9 (53)	14 (67)	10 (71)	0.525
Left	19 (37)	8 (47)	7 (33)	4 (29)	
Smoking status					
Current	4 (8)	2 (12)	2 (10)	0 (0)	0.224
Past	25 (48)	8 (47)	7 (33)	10 (71)	
Never	23 (44)	7 (41)	12 (57)	4 (29)	
Education					
Primary	10 (19)	3 (18)	2 (10)	5 (36)	0.270
Secondary	23 (44)	6 (35)	12 (57)	5 (36)	
Third level	19 (37)	8 (47)	7 (33)	4 (28)	
Variable	Mean ± SD (n = 52)	Mean ± SD (n = 17)	Mean ± SD (n = 21)	Mean ± SD (n = 14)	
Age	66 (8)	65 (7)	64 (9)	70 (8)	0.117
BMI, kg/m ²	26.1 (5.5)	25.5 (4.1)	27.1 (3.6)	25.2 (8.6)	0.562
CDVA (study eye)	99 (7)	99 (7)	99 (8)	98 (6)	0.868
Letter contrast sensitivity					
1.2 cpd	68.3 (46.4)	73.0 (49.1)	61.2 (41.3)	73.2 (52.3)	0.674
2.4 cpd	57.1 (41.2)	59.7 (45.3)	56.8 (40.5)	54.3 (41.4)	0.938
6.0 cpd	25.6 (14.8)	29.0 (14.9)	24.3 (14.0)	23.6 (16.0)	0.530
9.6 cpd	13.7 (8.6)	16.0 (9.1)	12.3 (7.3)	12.9 (9.7)	0.399
15.15 cpd	6.5 (4.9)	7.1 (4.5)	6.2 (4.8)	6.4 (5.7)	0.827
Macular pigment optical density					
0.25° eccentricity	0.50 (0.25)	0.50 (0.25)	0.50 (0.24)	0.47 (0.21)	0.925
0.5° eccentricity	0.39 (0.22)	0.38 (0.27)	0.41 (0.22)	0.36 (0.19)	0.797
1.0° eccentricity	0.26 (0.15)	0.27 (0.18)	0.27 (0.13)	0.24 (0.17)	0.851
1.75° eccentricity	0.14 (0.11)	0.16 (0.11)	0.15 (0.11)	0.11 (0.12)	0.554
Diet score* (n = 50)†	18.7 (11.2)	17.3 (10.9)	21.9 (12.7)	16.0 (8.4)	0.267

*A subject's weekly intake of carotenoid-rich foods was inputted into an L/Z screener to give a carotenoid-based diet score. Values are weighted for frequency of intake of the food and for bioavailability of L and Z within these foods (the range of scores on the L/Z screener is 0–75).

†Data were not available for 2 subjects.

BMI, body mass index; CDVA, corrected distance visual acuity; cpd, cycles per degree; SD, standard deviation.

Table 2. Mean (\pm SD) MPOD at Baseline and 12 Months

Eccentricity	Group 1			Group 2			Group 3		
	Baseline	12 Months	<i>P</i>	Baseline	12 Months	<i>P</i>	Baseline	12 Months	<i>P</i>
0.25°	0.50 \pm 0.25	0.59 \pm 0.30	0.077	0.50 \pm 0.25	0.60 \pm 0.21	0.005	0.46 \pm 0.21	0.59 \pm 0.20	0.010
0.5°	0.38 \pm 0.27	0.47 \pm 0.27	0.055	0.42 \pm 0.22	0.50 \pm 0.19	0.005	0.36 \pm 0.19	0.46 \pm 0.21	0.020
1°	0.27 \pm 0.18	0.34 \pm 0.16	0.083	0.27 \pm 0.13	0.34 \pm 0.17	0.005	0.24 \pm 0.17	0.33 \pm 0.16	0.019
1.75°	0.16 \pm 0.11	0.21 \pm 0.09	0.018	0.14 \pm 0.11	0.22 \pm 0.12	0.002	0.11 \pm 0.12	0.19 \pm 0.10	0.006

SD, standard deviation.

however, remained within their respective and normal reference ranges.

Discussion

The Meso-zeaxanthin Ocular Supplementation Trial is a randomized single-blind clinical trial that compares the effect of supplementation with three different macular carotenoid formulations on MPOD, visual performance, and AMD grade, over a period of 12 months, in subjects with early AMD.

The MPOD was significantly greater at 1 year than at baseline at all eccentricities for subjects in Groups 2 and 3. Although the observed augmentation in mean MPOD at 12 months did not reach statistical significance for subjects supplemented with high doses of L (Group 1) in the absence of MZ, except at 1.75° eccentricity, it should be noted that the mean increases observed for this group at other eccentricities were not dissimilar in magnitude to those observed for Groups 2 and 3.

The significant rise in MPOD across the spatial profile when all 3 macular carotenoids (Group 2) are included in the formulation, or when supplemented with 17 mg of MZ and small amounts of L and Z (Group 3), and especially the augmentation of MP centrally, is neither surprising nor counter-intuitive, given the known distribution of MP's individual constituent carotenoids.⁷ The inclusion of MZ in the formulation is likely to result in augmentation of MP centrally (demonstrated in Groups 2 and 3 here), as

this is the site of dominance of this carotenoid. In addition, the inclusion of L in the formulation (as in Groups 1, 2, and 3) will result in MP augmentation at the site of that carotenoid's natural dominance (1.75°), attested to by augmentation of MP at this locus in the high L (but no MZ) group (Group 1). It would seem, therefore, that supplementation with all three macular carotenoids results in the greatest augmentation of MPOD across its spatial profile, thereby putatively affording the greatest protection against AMD. Interestingly, *in vitro* work has concluded that the antioxidant capacity of the macular carotenoids is maximized when all three macular carotenoids are present.⁴²

It is unsurprising that there were demonstrable improvements in the CS after augmentation of MP, especially where such augmentation was demonstrated centrally, given the consequential enhancement of pre-receptor filtration of blue light and attenuation of the adverse effects of short-wavelength (blue) light scatter. This is particularly important for subjects with AMD because CS is an important measure of visual function in patients afflicted with the condition.⁵⁵ However, the inclusion of MZ in the formulation was required to achieve improvements both at low and high spatial frequencies.

The observation in this study that supplementation with high doses of L (in the absence of MZ) resulted in improved CS at low spatial frequencies only is consistent with the fact that visual function at low spatial frequencies will be mediated by slightly eccentric retinal loci. Of note, concentrations of L

Table 3. Mean (\pm SD) Letter CS Values at Baseline and at 12 Months

cpd	Group 1			Group 2			Group 3		
	Baseline	12 Months	<i>P</i>	Baseline	12 Months	<i>P</i>	Baseline	12 Months	<i>P</i>
1.2	73.0 \pm 49.1	91.8 \pm 48.5	0.021	61.2 \pm 41.3	91.9 \pm 53.6	0.014	73.2 \pm 52.3	92.2 \pm 55.0	0.081
2.4	59.7 \pm 45.3	86.7 \pm 54.2	0.006	56.8 \pm 40.5	77.8 \pm 51.6	0.008	54.3 \pm 41.4	86.2 \pm 52.1	0.002
6.0	29.0 \pm 14.9	38.1 \pm 26.7	0.098	24.3 \pm 14.0	30.9 \pm 18.8	0.058	23.6 \pm 16.0	42.2 \pm 25.9	0.002
9.6	16.0 \pm 9.1	16.4 \pm 9.0	0.939	12.3 \pm 7.3	17.5 \pm 12.3	0.066	12.9 \pm 9.7	20.1 \pm 11.9	0.016
15.15	7.1 \pm 4.5	7.8 \pm 5.5	0.408	6.2 \pm 4.8	7.8 \pm 6.4	0.189	6.4 \pm 5.7	8.7 \pm 5.8	0.005

Note: The statistical tests were based on log-transformed data. The *P* values reported are for the paired *t*-test (or the corresponding nonparametric test when the data distribution was non-normal).

cpd, cycles per degree; SD, standard deviation.

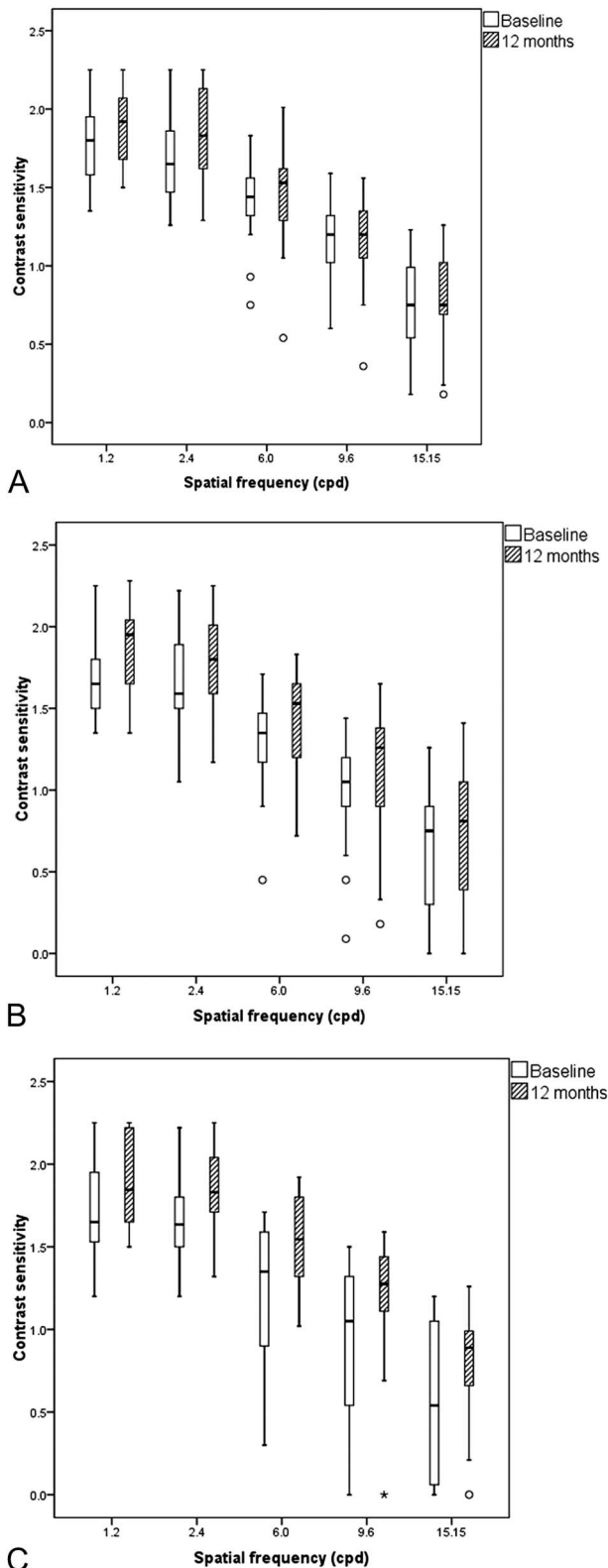


Fig 1. Letter CS at baseline and at 12 months, for Groups 1 (A), 2 (B), and 3 (C).

are higher in the peripheral macula, compared with the foveola.⁵⁶

Previous studies have investigated the impact of macular carotenoid supplementation on CS in subjects with AMD, with most of the studies reporting improvements in the CS after supplementation (with L and Z),^{36,37,57–59} although no study to date has tested a formulation containing MZ. For example, a study by Ma et al⁶⁰ has shown significant increases in CS at low spatial frequencies after supplementation with either 10 mg L, 20 mg L, or 10 mg L and 10 mg Z (combined), in subjects with early AMD, over a 48-week study period. These findings are in agreement with those reported in this study, which found demonstrable improvements in CS at high spatial frequencies, but only among subjects who were supplemented with a formulation containing MZ, and not among subjects supplementing with high doses of L alone.

This study has shown that, from a morphologic perspective, AMD remains stable for at least 12 months after supplementation with the macular carotenoids. However, the findings presented here must be interpreted with full appreciation of the study's principal weaknesses, and these include the small numbers of subjects involved, the study's short duration, and the absence of a placebo group. For the purposes of discussion, it is reasonable to compare our findings with the placebo group in the recently published Carotenoids in Age-Related Maculopathy study, which was a randomized, double-blind, placebo-controlled clinical trial of L (12 mg) and Z (0.6 mg) supplementation with co-antioxidants versus placebo in patients with early AMD.⁶¹ The study population of the Carotenoids in Age-Related Maculopathy study is comparable with that of our study, in terms of inclusion and exclusion criteria, methodology of AMD grading, and demographic and geographic considerations. However, in the Carotenoids in Age-Related Maculopathy study, at 12 months, 47.4% of eyes in the placebo arm (108 of 228 eyes) exhibited an increase of at least one grade (progression) along the AMD severity scale (data on file). Interestingly, in this study, only 27% of subjects (all of whom were supplementing with the macular carotenoids) showed progression by one or more steps at 12 months. Of course, a historical comparison such as this one should be interpreted with full appreciation of the fact that the natural course of AMD, particularly over a 12-month period, may not be clinically significant.

No discussion of our findings would be complete without reference to the recently published AREDS2 reports,^{62,63} where analysis of secondary outcomes indicated a benefit of supplementation with L and Z, in terms of AMD progression and preservation of

Table 4. Change in AMD Grade (11-Step Scale) Between Baseline and 12 Months

Group	n	-2 (%)	-1 (%)	0 (%)	+1 (%)	+2 (%)	+3 (%)	Significance
1	17	1 (5.9)	1 (5.9)	10 (58.8)	3 (17.6)	1 (5.9)	1 (5.9)	0.455
2	21	3 (14.3)	2 (9.5)	11 (52.4)	3 (14.3)	2 (9.5)	0	
3	14	2 (14.3)	5 (35.7)	4 (28.6)	2 (14.3)	1 (7.1)	0	
Total	52 (100%)	6 (11.5)	8 (15.4)	25 (48.1)	8 (15.4)	4 (7.7)	1 (1.9)	

Negative value, disease progression; positive value, disease regression; 0, no change in grade.

vision, especially in subjects with low dietary intake of those two carotenoids. Given that only two of the three macular carotenoids were used in AREDS2, our findings are rendered all the more clinically meaningful, if not somewhat provocative.

Of note, AREDS, published in 2001,⁶⁴ was criticized for non-inclusion of L and Z, and that omission prompted AREDS2. Our findings, however, suggest that the exclusion of MZ in AREDS2 represents a potential shortcoming of that study, especially given

Table 5. Clinical Pathology Variables Following Supplementation With the Macular Carotenoids Assessed at Baseline and at 12 months for Each of the Three Intervention Groups

Pathology Variable	Function of Test	Reference Range (Unit)*	Group 1 (n = 9)†		P
			Baseline	12 Months	
Sodium	Renal profile	135–145 (mmol/L)	139 ± 3	138 ± 3	0.312
Potassium	Renal profile	3.3–5.3 (mmol/L)	4.6 ± 0.3	4.7 ± 0.2	0.366
Chloride	Renal profile	98–107 (mmol/L)	104 ± 2	106 ± 2	0.073
Urea	Renal profile	2.5–7.7 (mmol/L)	7.2 ± 2.4	6.5 ± 1.4	0.174
Creatinine	Renal profile	40–90 (μmol/L)	81 ± 13	74 ± 10	0.086
Total protein	Liver profile	64–83 (g/L)	69 ± 3	68 ± 3	0.499
Albumin	Liver profile	37–52 (g/L)	41 ± 2	40 ± 3	0.444
Globulins	Liver profile	21–36 (g/L)	28 ± 4	28 ± 3	1.000
Total bilirubin	Liver profile	3.4–21.0 (μmol/L)	6.2 ± 2.0	7.8 ± 2.2	0.050
AAT	Liver profile	0–55 IU/L	23 ± 8	21 ± 8	0.426
ASA	Liver profile	5–36 IU/L	24 ± 3	24 ± 4	0.782
Alkaline phosphate	Liver profile	40–150 IU/L	79 ± 27	87 ± 31	0.013
GGT	Liver profile	9–36 IU/L	39 ± 40	40 ± 41	0.668
Cholesterol total	Lipid profile	<5.0 (mmol/L)	5.2 ± 1.0	5.2 ± 1.1	0.708
Triglycerides	Lipid profile	0.60–1.70 (mmol/L)	1.47 ± 0.61	1.34 ± 0.66	0.185
HDL	Lipid profile	1.00–1.55 (mmol/L)	1.51 ± 0.37	1.43 ± 0.31	0.063
Direct LDL	Lipid profile	<3.0 (mmol/L)	3.1 ± 1.0	3.2 ± 1.0	0.419
Calcium	Bone profile	2.10–2.60 (mmol/L)	2.31 ± 0.10	2.32 ± 0.14	0.661
Phosphate	Bone profile	0.80–1.56 (mmol/L)	1.13 ± 0.17	1.20 ± 0.24	0.292
Magnesium	Bone profile	0.65–1.10 (mmol/L)	0.99 ± 0.05	0.94 ± 0.09	0.159
Uric Acid	Bone profile	155–394 (μmol/L)	290 ± 54	280 ± 62	0.579
Glucose	Bone profile	3.1–6.1 (mmol/L)	5.3 ± 0.7	5.3 ± 1.2	0.910
HSRP	Inflammation marker	<5.0 (mg/L)	1.2 ± 0.5	1.6 ± 1.0	0.097
Full blood count					
Leukocyte count	Hematology	3.88–10.49 (10e9/L)	6.54 ± 2.00	5.88 ± 1.03	0.331
Erythrocyte count	Hematology	3.73–5.02 (10e12/L)	4.51 ± 0.42	4.36 ± 0.33	0.367
Hemoglobin	Hematology	11.3–15.2 (g/dL)	13.6 ± 1.1	13.6 ± 0.9	0.622
Hematocrit	Hematology	0.323–0.462 (L/L)	0.407 ± 0.032	0.405 ± 0.020	0.769
MCV	Hematology	83.1–99.1 (fL)	90.5 ± 3.1	93.1 ± 5.1	0.222
MCH	Hematology	28.3–33.9 (pg)	30.1 ± 1.2	31.3 ± 1.8	0.134
MCHC	Hematology	32.1–36.6 (g/dL)	33.3 ± 1.0	33.6 ± 0.8	0.357
Platelets	Hematology	164–382 (10e9/L)	332 ± 249	249 ± 123	0.196
Differential white cell count					
Neutrophils	Hematology	1.91–7.16 (10e9/L)	3.80 ± 1.27	3.44 ± 0.82	0.423
Lymphocytes	Hematology	1.01–3.13 (10e9/L)	1.82 ± 0.54	1.66 ± 0.39	0.309
Monocytes	Hematology	0.19–0.68 (10e9/L)	0.47 ± 0.18	0.40 ± 0.13	0.322

(continued on next page)

Table 5. (Continued)

Pathology Variable	Function of Test	Reference Range (Unit)*	Group 1 (n = 9)†		P
			Baseline	12 Months	
Eosinophils	Hematology	0.05–0.51 (10e9/L)	0.22 ± 0.05	0.18 ± 0.06	0.195
Basophils	Hematology	0.02–0.15 (10e9/L)	0.05 ± 0.03	0.05 ± 0.02	0.505
Large unstained cells	Hematology	0.00–0.30 (10e9/L)	0.17 ± 0.07	0.14 ± 0.03	0.222

Pathology Variable	Group 2 (n = 20)†			Group 3 (n = 12)†		
	Baseline	12 Months	P	Baseline	12 Months	P
Sodium	141 ± 3	138 ± 2	0.001	136 ± 3	137 ± 4	0.371
Potassium	4.6 ± 0.4	4.7 ± 0.4	0.475	4.7 ± 0.3	4.8 ± 0.2	0.709
Chloride	104 ± 3	104 ± 3	0.922	103 ± 4	103 ± 4	0.612
Urea	6.1 ± 1.1	6.6 ± 1.5	0.073	6.7 ± 1.5	6.0 ± 1.7	0.053
Creatinine	78 ± 14	77 ± 15	0.299	76 ± 19	75 ± 17	0.681
Total protein	71 ± 4	70 ± 3	0.415	70 ± 5	70 ± 5	0.558
Albumin	43 ± 2	42 ± 2	0.134	41 ± 2	42 ± 2	0.410
Globulins	28 ± 4	29 ± 3	0.737	29 ± 5	28 ± 4	0.272
Total bilirubin	9.1 ± 4.7	9.9 ± 5.3	0.293	8.0 ± 3.6	10.1 ± 4.0	0.001
AAT	22 ± 6	22 ± 6	0.752	19 ± 3	20 ± 5	0.279
ASA	22 ± 4	22 ± 4	0.903	21 ± 3	22 ± 4	0.083
Alkaline phosphate	78 ± 20	79 ± 20	0.501	76 ± 11	82 ± 17	0.114
GGT	27 ± 11	28 ± 14	0.395	27 ± 16	32 ± 23	0.075
Cholesterol total	4.7 ± 1.3	4.5 ± 0.9	0.231	4.8 ± 1.0	4.8 ± 0.9	1.000
Triglycerides	1.44 ± 0.49	1.39 ± 0.60	0.700	1.51 ± 1.31	1.29 ± 0.82	0.236
HDL	1.31 ± 0.33	1.24 ± 0.28	0.044	1.46 ± 0.47	1.46 ± 0.51	0.942
Direct LDL	2.8 ± 1.1	2.7 ± 0.8	0.317	2.8 ± 0.9	2.7 ± 0.9	0.671
Calcium	2.35 ± 0.07	2.35 ± 0.07	0.825	2.31 ± 0.06	2.40 ± 0.11	0.005
Phosphate	1.17 ± 0.17	1.19 ± 0.19	0.672	1.07 ± 0.25	1.10 ± 0.21	0.414
Magnesium	0.97 ± 0.08	0.98 ± 0.06	0.573	0.93 ± 0.12	0.94 ± 0.08	0.599
Uric Acid	315 ± 65	312 ± 66	0.724	305 ± 65	322 ± 89	0.260
Glucose	5.0 ± 0.6	5.0 ± 0.7	0.867	5.0 ± 0.9	5.1 ± 0.7	0.273
HSRP	2.2 ± 2.4	2.3 ± 2.1	0.864	4.0 ± 5.1	4.3 ± 6.7	0.728
Full blood count						
Leukocyte count	6.74 ± 1.53	6.81 ± 1.78	0.830	6.13 ± 1.56	5.95 ± 1.10	0.661
Erythrocyte count	4.50 ± 0.41	4.47 ± 0.39	0.377	4.44 ± 0.45	4.46 ± 0.45	0.858
Hemoglobin	13.7 ± 1.1	13.7 ± 1.2	0.596	13.6 ± 1.2	13.6 ± 1.1	0.969
Hematocrit	0.413 ± 0.031	0.413 ± 0.031	0.939	0.409 ± 0.031	0.412 ± 0.031	0.779
MCV	92.0 ± 4.0	92.6 ± 4.0	0.414	92.4 ± 3.7	92.6 ± 3.7	0.778
MCH	30.4 ± 1.3	30.8 ± 1.2	0.167	30.7 ± 1.2	30.5 ± 1.4	0.632
MCHC	33.1 ± 0.8	33.3 ± 1.2	0.523	33.2 ± 1.1	33.0 ± 1.0	0.468
Platelets	258 ± 88	250 ± 118	0.527	244 ± 46	254 ± 61	0.369
Neutrophils	4.09 ± 1.20	4.23 ± 1.38	0.580	3.92 ± 1.23	3.84 ± 1.21	0.809
Lymphocytes	1.81 ± 0.43	1.70 ± 0.42	0.128	1.42 ± 0.36	1.38 ± 0.39	0.704
Monocytes	0.43 ± 0.11	0.45 ± 0.16	0.495	0.41 ± 0.12	0.35 ± 0.09	0.132
Eosinophils	0.18 ± 0.09	0.23 ± 0.13	0.055	0.17 ± 0.10	0.17 ± 0.08	1.000
Basophils	0.05 ± 0.02	0.07 ± 0.04	0.063	0.04 ± 0.02	0.06 ± 0.03	0.042
Large unstained cells	0.18 ± 0.07	0.16 ± 0.05	0.122	0.16 ± 0.05	0.14 ± 0.02	0.177

*The normative reference ranges for this study were obtained from the insert kits for the instrumentation used by Biomnis Laboratories. Exceptions were the reference ranges for lipids (HDL, LDL, total cholesterol, and triglycerides), which were obtained from the European Guidelines on Cardiovascular Disease Prevention, (66) and for glucose, which were obtained from the World Health Organization (67).

†Total n ≠ 52 because data on pathology analysis was not available for all subjects at both baseline and 12 months.

AAT, alanine aminotransferase; ASA, aspartate aminotransferase; GGT, gamma glutamyl transpeptidase; HDL, high-density lipoprotein; LDL, low-density lipoprotein; HSCP, high sensitive reactive protein; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration; MCV, mean corpuscular volume. Values in bold are indicative of statistical significance at the 5% level.

that MP was measured in less than 2% of AREDS2 patients. Certainly, the opportunity to demonstrate further visual benefit by inclusion of all three of MP's constituent carotenoids was missed, and warrants fur-

ther study. In this vein, a head-to-head randomized, double-blind, controlled trial of the AREDS2 formula versus the AREDS2 formula fortified with MZ is currently underway.⁶⁵

Conclusion

Macular pigment can be augmented, and CS enhanced, in subjects with early AMD who receive supplemental macular carotenoids. A formulation containing all three macular carotenoids (L, Z, and MZ) may offer advantages over a formulation that does not contain MZ, in terms of improvements in CS and MP augmentation across its spatial profile, the latter putatively affording greater protection against (photo-) oxidative injury. The results of this study should inform and prompt a well-designed, controlled clinical trial of supplementation with L, Z, and MZ in subjects with AMD, where outcome measures should include visual function and disease progression.

Key words: lutein, zeaxanthin, *meso*-zeaxanthin, age-related macular degeneration, visual performance.

References

- Bressler NM. Age-related macular degeneration is the leading cause of blindness. *JAMA* 2004;291:1900–1901.
- Kelliher C, Kenny D, O'Brien C. Trends in blind registration in the adult population of the Republic of Ireland 1996–2003. *Br J Ophthalmol* 2006;90:367–371.
- Owen CG, Jarrar Z, Wormald R, et al. The estimated prevalence and incidence of late stage age related macular degeneration in the UK. *Br J Ophthalmol* 2012;96:752–756.
- Rosenfeld PJ, Rich RM, Lalwani GA. Ranibizumab: phase III clinical trial results. *Ophthalmol Clin North Am* 2006;19:361–372.
- Trieschmann M, van Kuijk FJ, Alexander R, et al. Macular pigment in the human retina: histological evaluation of localization and distribution. *Eye (Lond)* 2008;22:132–137.
- Hammond BR, Wooten BR, Snodderly DM. Individual variations in the spatial profile of human macular pigment. *J Opt Soc Am A Opt Image Sci Vis* 1997;14:1187–1196.
- Bone RA, Landrum JT, Hime GW, et al. Stereochemistry of the human macular carotenoids. *Invest Ophthalmol Vis Sci* 1993;34:2033–2040.
- Snodderly DM, Brown PK, Delori FC, Auran JD. The macular pigment. I. Absorbance spectra, localization, and discrimination from other yellow pigments in primate retinas. *Invest Ophthalmol Vis Sci* 1984;25:660–673.
- Tomany SC, Cruickshanks KJ, Klein R, et al. Sunlight and the 10-year incidence of age-related maculopathy: the Beaver Dam Eye study. *Arch Ophthalmol* 2004;122:750–757.
- Beatty S, Koh H, Phil M, et al. The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv Ophthalmol* 2000;45:115–134.
- Nolan JM, Stack J, O'Donovan O, Loane E, Beatty S. Risk factors for age-related maculopathy are associated with a relative lack of macular pigment. *Exp Eye Res* 2007;84:61–74.
- Seddon JM, Ajani UA, Sperduto RD, et al. Dietary carotenoids, vitamins A, C, and E, and advanced age-related macular degeneration. Eye Disease Case-Control Study Group. *JAMA* 1994;272:1413–1420.
- Mares-Perlman JA, Fisher AI, Klein R, et al. Lutein and zeaxanthin in the diet and serum and their relation to age-related maculopathy in the third national health and nutrition examination survey. *Am J Epidemiol* 2001;153:424–432.
- Snellen ELM, Verbeek ALM, Van Den Hoogen GWP, et al. Neovascular age-related macular degeneration and its relationship to antioxidant intake. *Acta Ophthalmol Scand* 2002;80:368–371.
- San Giovanni JP, Chew EY, Clemons TE, et al, AREDS Research Group. Dietary lipid intake and incident advanced age-related macular degeneration (AMD) in the Age-Related Eye Disease Study (AREDS). *Invest Ophthalmol Vis Sci* 2005;46:2382.
- Tan JS, Wang JJ, Flood V, et al. Dietary antioxidants and the long-term incidence of age-related macular degeneration the Blue Mountains Eye Study. *Ophthalmology* 2007;115:334–341.
- Antioxidant status and neovascular age-related macular degeneration. Eye Disease Case-Control Study Group. *Arch Ophthalmol* 1993;111:104–109.
- Delcourt C, Carriere I, Delage M, et al. Plasma lutein and zeaxanthin and other carotenoids as modifiable risk factors for age-related maculopathy and cataract: the POLA Study. *Invest Ophthalmol Vis Sci* 2006;47:2329–2335.
- Fletcher AE, Bentham GC, Agnew M, et al. Sunlight exposure, antioxidants, and age-related macular degeneration. *Arch Ophthalmol* 2008;126:1396–1403.
- Hammond BR, Johnson EJ, Russell RM, et al. Dietary modification of human macular pigment density. *Invest Ophthalmol Vis Sci* 1997;38:1795–1801.
- Johnson EJ, Hammond BR, Yeum KJ, et al. Relation among serum and tissue concentrations of lutein and zeaxanthin and macular pigment density. *Am J Clin Nutr* 2000;71:1555–1562.
- Bone RA, Landrum JT, Guerra LH, et al. Lutein and zeaxanthin dietary supplements raise macular pigment density and serum concentrations of these carotenoids in humans. *J Nutr* 2003;133:992–998.
- Trieschmann M, Beatty S, Nolan JM, et al. Changes in macular pigment optical density and serum concentrations of its constituent carotenoids following supplemental lutein and zeaxanthin: the LUNA study. *Exp Eye Res* 2007;84:718–728.
- Schalch W, Cohn W, Barker FM, et al. Xanthophyll accumulation in the human retina during supplementation with lutein or zeaxanthin—the LUXEA (LUtein Xanthophyll Eye Accumulation) study. *Arch Biochem Biophys* 2007;458:128–135.
- Richer S, Devenport J, Lang JC. LAST II: differential temporal responses of macular pigment optical density in patients with atrophic age-related macular degeneration to dietary supplementation with xanthophylls. *Optometry* 2007;78:213–219.
- Nolan JM, Loughman J, Akkali MC, et al. The impact of macular pigment augmentation on visual performance in normal subjects: COMPASS. *Vision Res* 2011;51:459–469.
- Sommerburg O, Keunen JE, Bird AC, van Kuijk FJ. Fruits and vegetables that are sources for lutein and zeaxanthin: the macular pigment in human eyes. *Br J Ophthalmol* 1998;82:907–910.
- Perry A, Rasmussen H, Johnson EJ. Xanthophyll (lutein, zeaxanthin) content in fruits, vegetables and corn and egg products. *J Food Compos Anal* 2009;22:9–15.
- Maoka T, Arai A, Shimizu M, Matsuno T. The first isolation of enantiomeric and meso-zeaxanthin in nature. *Comp Biochem Physiol B* 1986;83:121–124.
- Connolly EE, Beatty S, Thurnham DI, et al. Augmentation of macular pigment following supplementation with all three macular carotenoids: an exploratory study. *Curr Eye Res* 2010;35:335–351.
- Loughman J, Akkali MC, Beatty S, et al. The relationship between macular pigment and visual performance. *Vision Res* 2010;50:1249–1256.

32. Engles M, Wooten B, Hammond B. Macular pigment: a test of the acuity hypothesis. *Invest Ophthalmol Vis Sci* 2007;48:2922–2931.
33. Kvasakul J, Rodriguez-Carmona M, Edgar DF, et al. Supplementation with the carotenoids lutein or zeaxanthin improves human visual performance. *Ophthalmic Physiol Opt* 2006;26:362–371.
34. Stringham JM, Hammond BR Jr. The glare hypothesis of macular pigment function. *Optom Vis Sci* 2007;84:859–864.
35. Wooten BR, Hammond BR. Macular pigment: influences on visual acuity and visibility. *Prog Retin Eye Res* 2002;21:225–240.
36. Richer S, Stiles W, Statkute L, et al. Double-masked, placebo-controlled, randomized trial of lutein and antioxidant supplementation in the intervention of atrophic age-related macular degeneration: the Veterans LAST study (Lutein Antioxidant Supplementation Trial). *Optometry* 2004;75:216–230.
37. Richer SP, Stiles W, Graham-Hoffman K, et al. Randomized, double-blind, placebo-controlled study of zeaxanthin and visual function in patients with atrophic age-related macular degeneration: the Zeaxanthin and Visual Function Study (ZVF) FDA IND #78, 973. *Optometry* 2011;82:667–680.
38. Beatty S, Chakravarthy U, Nolan JM, et al. Secondary outcomes in a clinical trial of carotenoids with coantioxidants versus placebo in early age-related macular degeneration. *Ophthalmology* 2013;120:600–606.
39. Johnson EJ, Neuringer M, Russell RM, et al. Nutritional manipulation of primate retinas, III: effects of lutein or zeaxanthin supplementation on adipose tissue and retina of xanthophyll-free monkeys. *Invest Ophthalmol Vis Sci* 2005;46:692–702.
40. Bone RA, Landrum JT, Friedes LM, et al. Distribution of lutein and zeaxanthin stereoisomers in the human retina. *Exp Eye Res* 1997;64:211–218.
41. Bhosale P, Bernstein PS. Synergistic effects of zeaxanthin and its binding protein in the prevention of lipid membrane oxidation. *Biochim Biophys Acta* 2005;1740:116–121.
42. Li B, Ahmed F, Bernstein PS. Studies on the singlet oxygen scavenging mechanism of human macular pigment. *Arch Biochem Biophys* 2010;504:56–60.
43. Landrum JT, Bone RA. Lutein, zeaxanthin, and the macular pigment. *Arch Biochem Biophys* 2001;385:28–40.
44. Billsten HH, Bhosale P, Yemelyanov A, et al. Photophysical properties of xanthophylls in carotenoproteins from human retinas. *Photochem Photobiol* 2003;78:138–145.
45. Kirby ML, Beatty S, Loane E, et al. A central dip in the macular pigment spatial profile is associated with age and smoking. *Invest Ophthalmol Vis Sci* 2010;51:6722–6728.
46. Nolan JM, Akkali MC, Loughman J, et al. Macular carotenoid supplementation in subjects with atypical spatial profiles of macular pigment. *Exp Eye Res* 2012;101:9–15.
47. Wooten BR, Hammond BR, Land RI, Snodderly DM. A practical method for measuring macular pigment optical density. *Invest Ophthalmol Vis Sci* 1999;40:2481–2489.
48. Loane E, Stack J, Beatty S, Nolan JM. Measurement of macular pigment optical density using two different heterochromatic flicker photometers. *Curr Eye Res* 2007;32:555–564.
49. Stringham JM, Hammond BR. Macular pigment and visual performance under glare conditions. *Optom Vis Sci* 2008;85:82–88.
50. Klein R, Davis MD, Magli YL, et al. The Wisconsin age-related maculopathy grading system. *Ophthalmology* 1991;98:1128–1134.
51. Sparrow JM, Dickinson AJ, Duke AM. The Wisconsin age-related macular degeneration grading system: performance in an independent centre. *Ophthalmic Epidemiol* 1997;4:49–55.
52. Klein R, Klein BE, Knudtson MD, et al. Fifteen-year cumulative incidence of age-related macular degeneration: the Beaver Dam Eye Study. *Ophthalmology* 2007;114:253–262.
53. Connolly EE, Beatty S, Loughman J, et al. Supplementation with all three macular carotenoids: response, stability, and safety. *Invest Ophthalmol Vis Sci* 2011;52:9207–9217.
54. Cohen J. *Statistical Power Analysis for the Behavioral Sciences*. 2nd ed. Hillsdale, New Jersey: Lawrence Erlbaum Associates; 1988.
55. Stangos N, Voutas S, Topouzis F, Karampatakis V. Contrast sensitivity evaluation in eyes predisposed to age-related macular degeneration and presenting normal visual acuity. *Ophthalmologica* 1995;209:194–198.
56. Landrum JT, Bone RA, Moore LL, Gomez CM. Analysis of zeaxanthin distribution within individual human retinas. *Methods Enzymol* 1999;299:457–467.
57. Piermarocchi S, Saviano S, Parisi V, et al. Carotenoids in Age-related Maculopathy Italian Study (CARMIS): two-year results of a randomized study. *Eur J Ophthalmol* 2011;22:216–225.
58. Sasamoto Y, Gomi F, Sawa M, et al. Effect of 1-year lutein supplementation on macular pigment optical density and visual function. *Graefes Arch Clin Exp Ophthalmol* 2011;249:1847–1854.
59. Bartlett HE, Eperjesi F. Effect of lutein and antioxidant dietary supplementation on contrast sensitivity in age-related macular disease: a randomized controlled trial. *Eur J Clin Nutr* 2007;61:1121–1127.
60. Ma L, Yan SF, Huang YM, et al. Effect of lutein and zeaxanthin on macular pigment and visual function in patients with early age-related macular degeneration. *Ophthalmology* 2012;119:2290–2297.
61. Neelam K, Hogg RE, Stevenson MR, et al. Carotenoids and co-antioxidants in age-related maculopathy: design and methods. *Ophthalmic Epidemiol* 2008;15:389–401.
62. Chew EY, SanGiovanni JP, Ferris FL, et al. Lutein/zeaxanthin for the treatment of age-related cataract: AREDS2 randomized trial report no. 4. *JAMA Ophthalmol* 2013;131:843–850.
63. Chew EY, Clemons TE, SanGiovanni JP, et al. Secondary analyses of the effects of lutein/zeaxanthin on age-related macular degeneration progression: AREDS2 report no. 3. *JAMA Ophthalmol* 2013;132:142–149.
64. Kassoff A, Kassoff J, Buehler J, et al. A randomized, placebo-controlled, clinical trial of high-dose supplementation with vitamins C and E, beta carotene, and zinc for age-related macular degeneration and vision loss—AREDS report no. 8. *Arch Ophthalmol* 2001;119:1417–1436.
65. Akuffo KO, Beatty S, Stack J, et al. Central retinal enrichment supplementation trials: design and methodology. *Ophthalmic Epidemiol* 2014;21:111–123.
66. European Society of Cardiology. *European Guidelines on Cardiovascular Disease Prevention*; EJCPR 2007;14(suppl 2):S1–S113.
67. World Health Organization. *Definition and diagnosis of diabetes mellitus and intermediate hyperglycemia*. Geneva: WHO; 2006.